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Quick Step Tomato yellow leaf curl virus (TYLCV) ELISA Kit

Size: 96 T, 48T

Catalogue Number:QS0036PI

Assay Time: 60 minutes

Store all reagents at 2-8°C/-20°C

Validity Period: 2-8°C for six months, -20°C for one year. Avoid repeated thaw cycles.

For samples:In serum, plasma, culture media or any biological fluid.

FOR RESEARCH USE ONLY !

NOT FOR THERAPEUTIC OR DIAGNOSTIC APPLICATIONS !

PLEASE READ THROUGH ENTIRE PROCEDURE BEFORE BEGINNING !

Quick Step Tomato yellow leaf curl virus (TYLCV)

ELISA Kit

FOR RESEARCH USE ONLY

Purpose

Our Quick Step Tomato yellow leaf curl virus (TYLCV) ELISA Kit is to for the qualitative determination of TYLCV in Tomato serum, plasma, culture media or any biological fluid.

Principle

The ELISA is based on the the qualitative enzyme immunoassay technique. The Microplate provided in this kit has been pre-coated with an antibody specific to TYLCV, make it to solid-phase antibody. Samples are added to the Microplate wells and combined to the specific antibody. Then a Horseradish Peroxidase (HRP)-conjugated antibody specific for TYLCV is added to each Microplate well and incubated, so the antibody-antigen-Enzyme labeled antibody complex is formed. Following a wash to remove any unbound reagent, then the TMB substrate solution is added to each well. Only those wells that contain TYLCV and HRP conjugated TYLCV antibody will appear blue in color and then turn yellow after the addition of the stop solution. The optical density (OD) is measured spectrophotometrically at a wavelength of 450 nm. The qualitative determination of TYLCV is determined by comparing with the CUT OFF value.

Materials provided with the kit

	Materials provided with the kit	96 determinations	48 determinations
1	User manual	1	1
2	Closure plate membrane	2	2
3	Sealed bags	1	1

4	Microelisa stripplate	1	1
5	Negative control	0.5ml×1 bottle	0.5ml×1 bottle
6	Positive control	0.5ml×1 bottle	0.5ml×1 bottle
7	HRP-Conjugate reagent	6ml×1 bottle	3ml×1 bottle
8	Sample diluent	6ml×1 bottle	3ml×1 bottle
9	Chromogen Solution A	6ml×1 bottle	3ml×1 bottle
10	Chromogen Solution B	6ml×1 bottle	3ml×1 bottle
11	Stop Solution	6ml×1 bottle	3ml×1 bottle
12	wash solution	20ml (30X)×1bottle	20ml (20X)×1bottle

Sample preparation

1. Serum preparation

After collection of the whole blood, allow the blood to clot by leaving it undisturbed at room temperature. This usually takes 10-20 minutes. Remove the clot by centrifuging at 2,000-3,000 rpm for 20 minutes. If precipitates appear during reservation, the sample should be centrifugated again.

2. Plasma preparation

Collect the whole blood into tubes with anticoagulant (EDTA or citrate). After incubated at room temperature for 10-20 minutes, tubes are centrifugated for 20 min at 2,000-3,000 rpm. Collect the supernatant carefully as plasma samples. If precipitates appear during reservation, the sample should be centrifugated again.

3. Urine samples

Collect urine into aseptic tubes. Collect the supernatant carefully after centrifuging for 20 min at 2,000-3,000 rpm. If precipitates appear during reservation, the sample should be centrifugated again. The preparation procedure of cerebrospinal fluid and pleuroperitoneal fluid is the same as that of urine sample.

4. Cell samples

If you want to detect the secretions of cells, collect culture supernatant into aseptic tubes. Collect the supernatant carefully after centrifuging for 20 min at 2,000-3,000 rpm. If you want to detect intracellular components, dilute the cells to 1X100/ml with PBS (pH 7.2-7.4). The cells were destroyed to release intracellular components by repeated freezing and thawing. Collect the supernatant carefully after centrifuging for 20 min at 2,000-3,000 rpm.

If precipitates appear during reservation, the sample should be centrifuged again.

5. Tissue samples

Tissue samples are cut, weighed, frozen in liquid nitrogen and stored at -80°C for future use. The tissue samples were homogenized after adding PBS (pH 7.4). Samples should be operated at 4°C . Collect the supernatant carefully after centrifuging for 20 min at 2,000-3,000 rpm. Aliquot the supernatant for ELISA assay and future use.

Notes:

1. Sample extraction and ELISA assay should be performed as soon as possible after sample collection. The samples should be extracted according to the relevant literature. If ELISA assay can not be performed immediately, samples can be stored at -20°C . Repeated freeze-thaw cycles should be avoided.
2. Our kits can not be used for samples with NaN_3 which can inhibit the activity of HRP.

Procedure

1. In the Microplate, number the corresponding micropores of the sample in sequence, leave two wells as negative control, two wells as positive control and one empty well as blank control. (blank control hole dont add samples and HRP-Conjugate reagent, the rest step operation are same)
2. **Add sample:** Set blank wells separately (blank comparison wells don't add sample and HRP-Conjugate reagent, other each step operation is same). testing sample well. add Sample dilution $40\mu\text{l}$ to testing sample well, then add testing sample $10\mu\text{l}$ (sample final dilution is 5-fold), add sample to wells, don't touch the well wall as far as possible, and Gently mix.
3. In the Microplate, number the corresponding micropores of the sample in sequence, leave two wells as negative control, two wells as positive control and one empty well as blank control. (blank control hole dont add samples and HRP-Conjugate reagent, the rest step operation are same)
4. **Add enzyme:** Add HRP-Conjugate reagent $50\mu\text{l}$ to each well, except blank well.

5. **Incubate:** After closing plate with Closure plate membrane ,incubate for **30 min** at 37°C.
6. **Configure liquid:** 30-fold (or 20-fold)wash solution diluted 30-fold (or 20-fold) with distilled water and reserve.
7. **Washing:** Uncover Closure plate membrane, discard Liquid, dry by swing, add washing buffer to every well, still for 30s then drain, repeat 5 times, dry by pat.
8. **Color:** Add Chromogen Solution A 50ul and Chromogen Solution B to each well, evade the light preservation for **10 min** at 37°C
9. **Stop the reaction:** Add Stop Solution50μl to each well, Stop the reaction(the blue color change to yellow color).
10. **Assay:** take blank well as zero, Read absorbance at 450nm after Adding Stop Solution and within **15min**.

Notes:

1. Store the kit at upon receipt.The kit should be equilibrated to room temperature before the assay. Remove any unneeded strips from Tomato TYLCV antibody-Coated plate, reseal them in zip-lock foil and keep at 2-8°C/-20° C.
2. Precipitates may appear in concentrated washing buffer. Please heat the buffer to dissolve all the precipitates, which will not affect the results.
3. Accurate pipette should be used to avoid experimental error. Samples should be added to the Microplate in less than 5 minutes. If a large number of samples are included, multiple channel pipette is recommended.
4. Standard curve should be included in every assay. Replicate wells are recommended. If the OD value of the sample is greater than the first well of standards, please dilute the sample (n times) before test. When calculating the original TYLCV concentration, please multiply the total dilution factor (XnX5).
5. In order to avoid cross-contamination, Microplate sealers are for one-time use only.
6. Please keep Substrate away from light.
7. All the operation should be accordance with the manufacturer's instructions strictly. The results determined by the Microtiter Plate Reader.
8. All the samples, washing buffer and wastes should be treated as infectious agents.

9. Reagents from different lots should not be mixed.

Determine the result

Test effectiveness: the average value of positive control ≥ 1.00 ; The average value of negative control ≤ 0.10 .

The critical value (CUT OFF) calculation: critical value = the average value of negative control + 0.15

Negative judgement: if the OD value < CUT OFF, the sample is Tomato TYLCV negative.

Positive judgement: if the OD value \geq CUT OFF, the sample is Tomato TYLCV positive.

Storage and validity

1. Storage: 2-8°C / -20° C.
2. Duration: 2-8°C for six months, -20° C for one year. Avoid repeated thaw cycles.